

Total Synthesis of (+)-Iresin

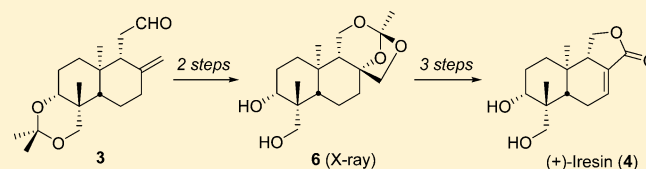
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Supporting Information

ABSTRACT: The first asymmetric total synthesis of (+)-iresin (**4**), an historically important *ent*-Drimane sesquiterpene lactone, was realized from aldehyde **3** via cyclic orthoester **6** in 5 steps. Notable transformations in this synthesis include a tandem trifluoroperacetic acid (TFPAA)-mediated Baeyer–Villiger oxidation–olefin epoxidation–epoxy ester cyclization, regioselective Burgess dehydration, and regioselective Fétizon oxidative lactonization.



Drimane (known also as *Iresane*) sesquiterpenoids constitute a group of widely occurring terpene natural products possessing a characteristic *bicyclofarnesol* skeleton (Figure 1).¹ Thus, the Drimane was once regarded as the missing biogenetic link between the lower and higher terpenoids.² Oxygenated Drimane sesquiterpenoids exhibit a wide range of significant biological activities,^{1,3} such as antifeedant, insecticidal, and cytotoxic, and have stimulated a great deal of attention on synthetic development.⁴ (+)-Iresin (**4**), a unique *ent*-Drimane sesquiterpene lactone, was first isolated by Djerassi and co-workers⁵ in 1954 from the Mexican herb plant *Iresine celosioides* (known as “herb of the mayas”) and characterized⁶ in 1958 by extensive chemical and spectroscopic methods and X-ray crystallographic study as the tricyclic dihydroxy lactone **4**. Full NMR spectroscopic assignments of **4** appeared in 2005.⁷ The sole documented classical synthetic study on **4** by Pelletier and Prabhakar⁸ in 1968 led to the total synthesis of isoresin in a racemic form.

In a previous publication of this serial study,⁹ we have reported the total synthesis of (–)-andrographolide (**1**) from a homoiido allylsilane epoxide **2** by a conformationally directed biomimetic cationic cyclization to form aldehyde **3** as a pivotal intermediate (Figure 1). We describe in this Note the first asymmetric total synthesis of (+)-iresin (**4**) from the same intermediate **3**. The conversion of aldehyde **3** to lactone **4** requires the elaboration of an unsaturated γ -lactone function, which would be achieved via oxidative cleavage of the formyl carbon and regioselective olefin formation and subsequent lactonization.

The synthetic route to **4** commenced with aldehyde **3** and is outlined in Schemes 1 and 2. To facilitate the Baeyer–Villiger oxidation,¹⁰ **3** was converted to the corresponding methyl ketone derivative **5** via a standard methylation–oxidation procedure. After screening of peroxy acids, freshly prepared trifluoroperacetic acid (TFPAA, ca. 0.2 M) was found to be the optimal oxidant in the presence of excess Na₂HPO₄ in CH₂Cl₂ at 0 °C. Keto olefin **5** was thus transformed to the cyclic orthoester **6**¹¹ as the major product (53%), along with the

corresponding hydrolytic product monoacetates **7a** and **7b** (17%).^{11a,e} The stereostructure of **6** was confirmed by a single-crystal X-ray diffraction analysis.¹² It is worthwhile to note that not only the Baeyer–Villiger oxygenation of the keto function occurred smoothly but also the olefin epoxidation and subsequent epoxide-opening–cyclization took place, leading to the cyclic orthoester **6**. The acetonide function of **5** was also unexpectedly cleaved under these reaction conditions. The unusual one-pot formation of cyclic orthoester **6** could be understood via the intramolecular rearrangement of an epoxy ester intermediate **5a**, as depicted in Scheme 1, based on serial mechanistic studies by Giner et al.^{11c–e} Acidic hydrolysis of orthoester **6** afforded a product mixture of monoacetates **7a** and **7b**,^{11a,e} which was subjected to acetylation to give the tetrakis-acetate **8** in 68% yield from **6** (Scheme 2). Standard dehydration of **8** employing various dehydrating agents (i.e., MsCl, SOCl₂, or POCl₃) in the presence of pyridine or Et₃N led to, after deacetylation, regioisomer **10** predominately (2.6–4.1:1) in good yield. In contrast, with Burgess reagent (3 equiv),¹³ dehydration of **8** in warm benzene (50 °C) gave, after deacetylation, the desired regioisomer **9** as the major product (4.8:1) in 70% overall yield.¹⁴

With **9** and **10** in hand, we selected the Fétizon reagent¹⁵ for the final oxidative lactonization (Scheme 2). To our delight, oxidation of **9** with freshly prepared Fétizon reagent in refluxing benzene produced the desired target lactone (+)-iresin (**4**) chemoselectively in 78% yield, along with aldehyde **11** in 14% yield. Reduction of **11** with NaBH₄ at 0 °C in methanol gave (+)-iresin (**4**) in 86% yield. Synthetic **4** exhibits identical spectroscopic data with those of reported (+)-iresin.^{5,6} Further structural confirmation of synthetic **4** was provided by the X-ray crystallographic analysis of the bis-*para*-bromobenzoate derivative **14**.^{6e,16} Interestingly, oxidation of **10** with excess Fétizon reagent in refluxing benzene furnished lactone aldehyde **12** chemoselectively in 91% yield, which was reduced with

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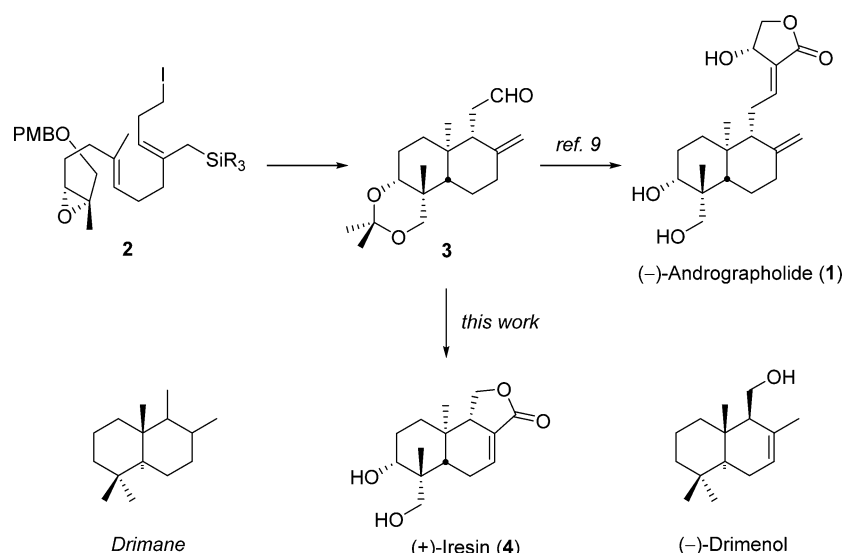
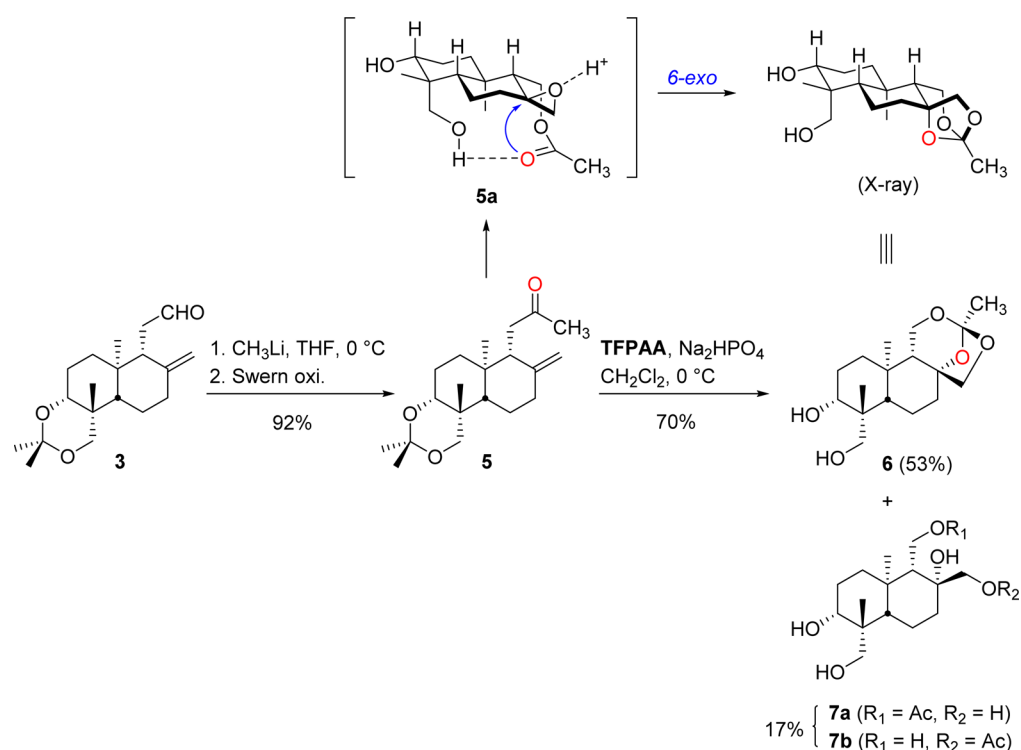


Figure 1. Structure of (+)-iresin and previous total synthesis of (–)-andrographolide (1) from homoiodo allylsilane epoxide 2 via aldehyde intermediate 3.

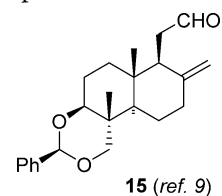
Scheme 1. Synthesis of Orthoester 6 from Aldehyde 3



NaBH_4 at 0 °C to give lactone 13 in 92% yield. The structure of synthetic 13, known as isoiresin,^{8,17} was further confirmed by a single-crystal X-ray crystallographic analysis.¹⁸

In summary, the first asymmetric total syntheses of (+)-iresin (4) and (–)-isoiresin (13) were achieved from readily accessible aldehyde 3 in 5 and 6 steps, respectively. Notable transformations include the peroxidation of keto olefin 5 with TFPAA, leading to cyclic orthoester 6 via a tandem Baeyer–Villiger oxidation–olefin epoxidation–epoxy ester cyclization, regioselective dehydration of 8 with Burgess reagent, as well as the regioselective Fétizon oxidative γ -lactonization of 9 and 10. The biomimetic synthetic approach¹⁹ demonstrated here for the *ent*-Drimanes would be equally effective for other Drimane

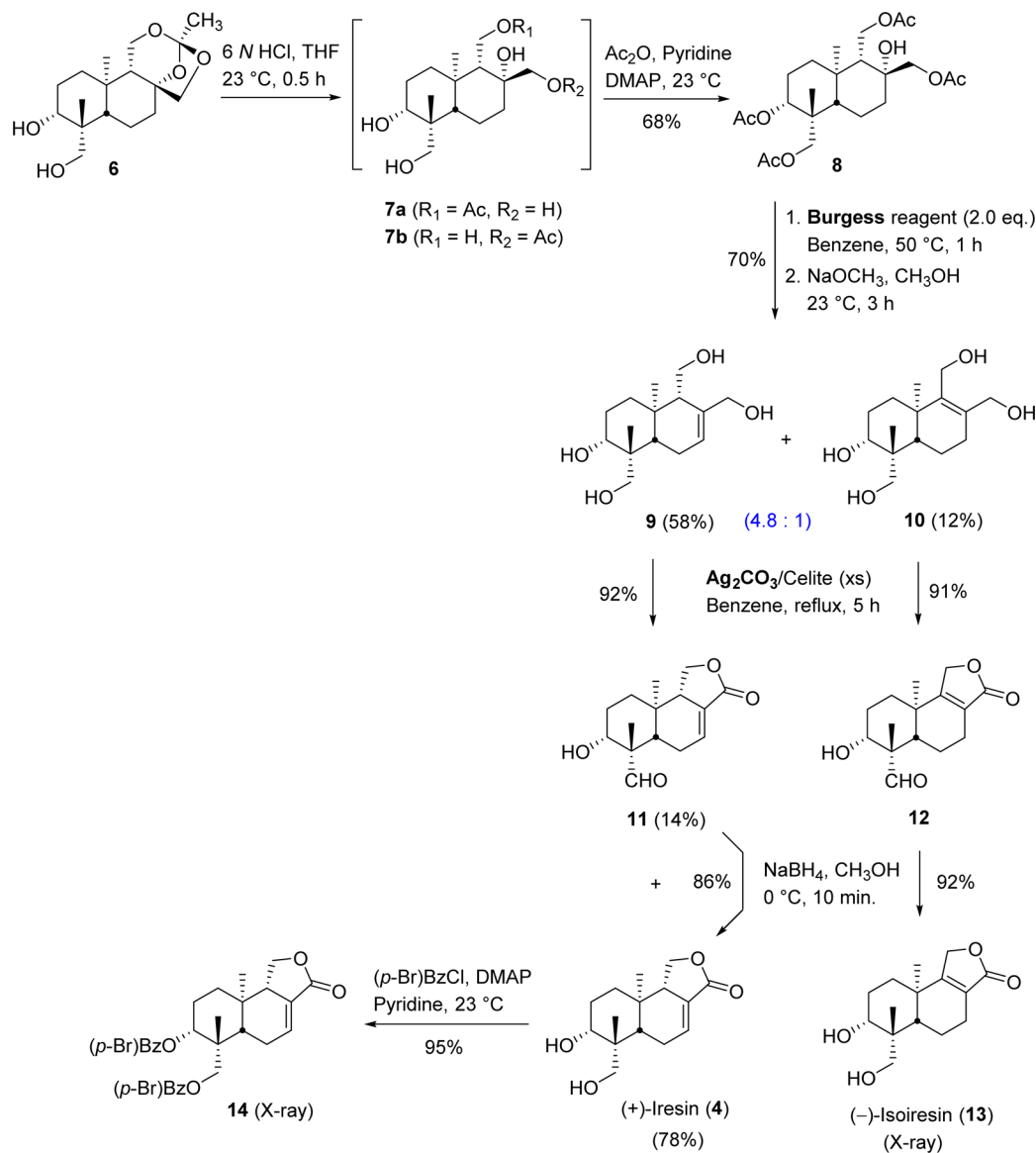
sesquiterpenoids, i.e., starting from synthetically readily accessible pseudo-antipodal intermediate 15.⁹



EXPERIMENTAL SECTION

General. For product purification by flash column chromatography, silica gel (200–300 mesh) and petroleum ether (bp. 60–90 °C) were

Scheme 2. Total Syntheses of (+)-Iresin (4) and (-)-Isoiresin (13)



used unless otherwise noted. All solvents were purified and dried by standard techniques, and distilled prior to use. Other commercially available reagents were used as received without further purification unless otherwise indicated. All organic extracts were dried over anhydrous sodium sulfate or magnesium sulfate. All moisture-sensitive reactions were carried out under an atmosphere of nitrogen in glassware that had been flame-dried under vacuum. ^1H and ^{13}C NMR spectra were recorded on a 400 MHz spectrometer with TMS as an internal reference and CDCl_3 as solvent, unless otherwise indicated. IR spectra were recorded on an FT-IR spectrometer as liquid film or KBr pellet. HRMS were acquired on an FT-ICR spectrometer. Melting points were measured on a hot stage and were uncorrected.

1-((4a*R*,6a*S*,7*R*,10a*S*,10b*R*)-3,3,6a,10b-Tetramethyl-8-methylenedecahydro-1*H*-naphtho[2,1-*d*][1,3]dioxin-7-yl)-propan-2-one (5). To a stirred solution of aldehyde (1.20 g, 3.92 mmol) in dry Et_2O (20 mL) was added 2.5 mL of CH_3Li (1.6 M in diethyl ether, 4.0 mmol) dropwise at $0\text{ }^\circ\text{C}$ under N_2 . After being stirred for 10 min, the reaction mixture was quenched with water (2.0 mL). The ethereal layer was washed with water and brine, dried over Na_2SO_4 , and concentrated in vacuo. The residue was used in the next step without further purification. DMSO (610 mg, 7.81 mmol) in CH_2Cl_2 (4.0 mL) was added dropwise to a solution of oxalic dichloride (0.33 mL, 3.87 mmol) in CH_2Cl_2 (4.0 mL) at $-78\text{ }^\circ\text{C}$

under N_2 . After stirring for 15 min, the solution of crude alcohol in CH_2Cl_2 (6.0 mL) was added via syringe, and the reaction was stirred an additional 1 h at $-78\text{ }^\circ\text{C}$. Et_3N (2.2 mL, 15.7 mmol) was added, and the reaction was stirred for 10 min at $-78\text{ }^\circ\text{C}$. The bath was removed, and the resulting mixture was stirred for 20 min at room temperature. The mixture was partitioned between H_2O (15 mL) and CH_2Cl_2 (30 mL), and the layers were separated. The aqueous layer was extracted with CH_2Cl_2 ($2 \times 15\text{ mL}$), and the combined organic layers were dried (Na_2SO_4), filtered, and concentrated under reduced pressure. The residue was purified by flash silica gel column chromatography (petroleum ether/ EtOAc = 8:1) to give compound 5 (1.15 g, 92% from 3) as a colorless oil: R_f = 0.51 (petroleum ether/ EtOAc = 8:1); $[\alpha]_D^{20}$ +20 (c 1.0, CHCl_3); IR (film): ν_{max} 3078, 2936, 2891, 1717, 1644, 1374, 1227, 1095, 885 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 4.77 (s, 1H), 4.37 (s, 1H), 3.97 (d, J = 11.6 Hz, 1H), 3.50 (dd, J = 8.4, 4.0 Hz, 1H), 3.18 (d, J = 11.6 Hz, 1H), 2.68 (dd, J = 17.2, 10.4 Hz, 1H), 2.45–2.38 (m, 3H), 2.17 (s, 3H), 2.08 (td, J = 12.0, 4.8 Hz, 1H), 2.02–1.96 (m, 1H), 1.81–1.71 (m, 2H), 1.58 (ddd, J = 23.6, 7.6, 5.6 Hz, 1H), 1.41 (s, 3H), 1.37 (s, 3H), 1.35–1.26 (m, 3H), 1.21 (s, 3H), 0.91 (s, 3H) ppm; ^{13}C NMR (100 MHz, CDCl_3): δ 208.3, 148.3, 107.2, 99.0, 76.4, 63.9, 52.0, 51.1, 40.1, 37.8, 37.7, 37.3, 34.4, 30.1, 27.2, 26.1, 25.3, 25.0, 23.0, 16.5 ppm; HRMS (ESI): calcd for $\text{C}_{20}\text{H}_{33}\text{O}_3$ $[\text{M} + \text{H}]^+$ 321.2424; found 321.2428.

(**3S,5aR,7aS,8R,9R,11aR,11bR**)-8-(Hydroxymethyl)-3,8,11a-trimethyldecahydro-1*H*-3,5a-epoxynaphtho[1,2-*e*][1,3]-dioxepin-9-ol (**6**). To a stirred solution of 50% aq H₂O₂ (112 mg, 1.65 mmol) in CH₂Cl₂ (8.0 mL) was added TFAA (0.28 mL, 1.99 mmol) at 0 °C. After being stirred for 25 min, powdered Na₂HPO₄ (1.13 g, 7.96 mmol) was added to the mixture; then the solution of **5** (106 mg, 0.33 mmol) in CH₂Cl₂ (2.0 mL) was added dropwise over 5 min. After stirring for 25 min at 0 °C, the reaction mixture was quenched with 5 mL of saturated aqueous Na₂S₂O₃. The resulting mixture was extracted with ethyl acetate (3 × 30 mL), and the combined organic extracts were washed with water and brine, and dried over anhydrous Na₂SO₄. The solvent was evaporated under reduced pressure, and the residue was purified by silica gel column chromatography eluting with 1:2 petroleum ether/EtOAc to give **6** as white solids (55 mg, 53%) and the mixture of monoacetate products (**7a** and **7b**, 19 mg, 17%). Compound **6**: *R*_f = 0.39 (petroleum ether/EtOAc = 1:2); mp 118–121 °C; [α]_D²⁰ +6 (c 1, CHCl₃); IR (KBr): ν_{max} 3365, 2938, 2886, 1463, 1402, 1301, 1134, 863 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 4.26 (d, *J* = 11.2 Hz, 1H), 4.03 (dd, *J* = 12.4, 5.6 Hz, 1H), 3.87 (d, *J* = 7.2 Hz, 1H), 3.82 (d, *J* = 12.4 Hz, 1H), 3.43–3.35 (m, 3H), 2.82 (d, *J* = 7.6 Hz, 1H), 2.73 (s, 1H), 1.92–1.84 (m, 3H), 1.78–1.67 (m, 2H), 1.62 (dd, *J* = 12.8, 3.2 Hz, 1H), 1.58–1.49 (m, 1H), 1.55 (s, 3H), 1.26 (s, 3H), 1.14 (s, 3H), 0.94–0.89 (m, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 119.7, 80.3, 79.8, 74.8, 63.9, 58.7, 53.7, 50.2, 42.4, 37.4, 36.9, 33.6, 27.2, 22.7, 22.3, 18.5, 16.7 ppm; HRMS (ESI): calcd for C₁₇H₂₈O₅ [M + H]⁺ 313.2010; found 313.2003. X-ray crystallographic data of **6**: C₁₇H₂₈O₅, triclinic, space group: *P*1, *a* = 6.108 (2) Å, *b* = 7.159 (3) Å, *c* = 20.206 (7) Å, α = 97.983(18)°, β = 91.80 (2)°, γ = 111.632(18)°, *Z* = 1, *d*_{calcd} = 1.281 g/cm³, *R*₁(*I* > 2σ(*I*)) = 0.0451, *wR*₂ = 0.1065. Compound **7a**: ¹H NMR (400 MHz, CD₃OD): δ 4.15 (d, *J* = 11.2 Hz, 1H), 4.06 (d, *J* = 11.2 Hz, 1H), 3.95 (d, *J* = 10.8 Hz, 1H), 3.91 (dd, *J* = 12.0, 4.0 Hz, 1H), 3.76 (dd, *J* = 11.6, 2.8 Hz, 1H), 3.40–3.34 (m, 2H), 2.05 (s, 3H, CH₃CO), 1.95 (d, *J* = 12.8 Hz, 1H), 1.85–1.70 (m, 3H), 1.65–1.63 (m, 2H), 1.54–1.49 (m, 1H), 1.21 (s, 3H), 1.45–1.37 (m, 2H), 1.06 (s, 3H), 0.97 (t, *J* = 7.2 Hz, 1H); ¹³C NMR (100 MHz, CD₃OD): δ 173.0, 81.2, 74.6, 71.8, 65.2, 59.5, 56.5, 56.4, 43.8, 39.0, 38.7, 38.3, 28.4, 23.6, 21.0, 19.0, 17.8 ppm. Compound **7b**: ¹H NMR (400 MHz, CD₃OD): δ 4.25–4.24 (m, 2H), 4.14 (d, *J* = 11.2 Hz, 1H), 3.43 (d, *J* = 10.8 Hz, 1H), 3.40–3.34 (m, 2H), 3.20 (d, *J* = 10.8 Hz, 1H), 2.01 (s, 3H, CH₃CO), 1.89 (dt, *J* = 13.2, 3.2 Hz, 1H), 1.82–1.79 (m, 1H), 1.73–1.63 (m, 6H), 1.40 (t, *J* = 3.6 Hz, 1H), 1.22 (s, 4H), 0.98 (s, 3H); ¹³C NMR (100 MHz, CD₃OD): δ 173.1, 81.1, 74.7, 69.8, 65.2, 63.0, 56.5, 53.6, 43.8, 39.0, 38.6, 38.1, 28.3, 23.5, 21.3, 19.0, 17.4 ppm.

(**1R,2R,4aS,5R,6R,8aR**)-6-Acetoxy-2-hydroxy-5,8a-dimethyldecahydronaphthalene-1,2,5-triyltris(methylene) Triacetate (**8**). To a stirred solution of **6** (260 mg, 0.83 mmol) in THF (4.0 mL) was added 6 N aq HCl (2.0 mL). After 30 min, the reaction mixture was diluted with water (5.0 mL) and extracted with ethyl acetate (4 × 20 mL). The combined organic extracts were washed with water and brine, dried over Na₂SO₄, and concentrated in vacuo to give a solid mixture of monoacetate products **7a** and **7b**. To a solution of products **7a** and **7b** in pyridine (2.0 mL) were added Ac₂O (0.50 mL, 5.3 mmol) and DMAP (9 mg, 0.074 mmol). After stirring overnight at room temperature, the reaction mixture was extracted with ethyl acetate, then washed with saturated NaHCO₃ solution, water and brine, dried over MgSO₄, and concentrated in vacuo. The residue was purified by flash silica gel column chromatography (petroleum ether/EtOAc = 3:1) to give compound **8** (257 mg, 68% from **6**, 80% from mixture of **7a** and **7b**) as a colorless oil. Compound **8**: *R*_f = 0.50 (petroleum ether/EtOAc = 1:1); [α]_D²⁰ –27 (c 1.0, CHCl₃); IR (film): ν_{max} 3498, 2949, 1738, 1441, 1372, 1246, 1037, 985 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 4.59 (dd, *J* = 9.6, 7.2 Hz, 1H), 4.36 (d, *J* = 12.0 Hz, 1H), 4.28 (d, *J* = 4.0 Hz, 2H), 4.17 (d, *J* = 12.0 Hz, 1H), 3.98 (d, *J* = 11.2 Hz, 1H), 3.95 (d, *J* = 11.6 Hz, 1H), 2.09 (s, 3H), 2.06 (s, 3H), 2.04 (s, 3H), 2.03 (s, 3H), 1.97 (d, *J* = 13.2 Hz, 1H), 1.85–1.80 (m, 2H), 1.76–1.69 (m, 4H), 1.49–1.41 (m, 1H), 1.34–1.25 (m, 2H), 1.07 (d, *J* = 12.0 Hz, 1H), 1.04 (s, 3H), 1.03 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 170.9, 170.9, 170.6, 170.5, 79.7, 72.6, 70.9, 65.0, 61.3, 55.1, 53.3, 41.1, 37.6, 37.5, 23.3, 22.7, 21.2, 21.1, 20.9, 18.1, 16.2 ppm;

HRMS (ESI): calcd for C₂₃H₄₀NO₉ [M + NH₄]⁺ 474.2698; found 474.2704.

((**1S,4aS,5R,6R,8aR**)-6-Hydroxy-5,8a-dimethyl-1,4,4a,5,6,7,8,8a-octahydronaphthalene-1,2,5-triyl)-trimethanol (**9**) and ((**4aS,5R,6R,8aR**)-6-Hydroxy-5,8a-dimethyl-3,4,4a,5,6,7,8,8a-octahydronaphthalene-1,2,5-triyl)-trimethanol (**10**). Compound **8** (250 mg, 0.55 mmol) in 2 mL of benzene was added dropwise to a solution of Burgess reagent (390 mg, 1.64 mmol); Burgess reagent was prepared according to the literature methods^{13d} in 3 mL of benzene at 50 °C under N₂. After being stirred for 1 h at 50 °C, the reaction mixture was diluted with water (5.0 mL) and extracted with diethyl ether (3 × 10 mL). The combined organic layers was washed with water and brine, dried over Na₂SO₄, and concentrated in vacuo to give a liquid residue. The residue was dissolved in dry CH₃OH (5.0 mL), and NaOCH₃ (148 mg, 2.74 mmol) was added. The reaction mixture was stirred for 3 h, then filtered through a pad of Celite, eluting with CHCl₃/CH₃OH (10:1). The resulting filtrate was concentrated under reduced pressure and purified by flash silica gel column chromatography CHCl₃/CH₃OH (25:1) to give 86 mg (58%) of **9** as white solids and 18 mg (12%) of **10** as white amorphous solids. Compound **9**: *R*_f = 0.46 (CHCl₃/CH₃OH = 10:1); mp 160–162 °C; [α]_D²⁰ –3 (c 1.0, CH₃OH); IR (KBr): ν_{max} 3327, 2930, 2858, 1707, 1448, 1363, 1024, 994 cm⁻¹; ¹H NMR (400 MHz, CD₃OD): δ 5.77 (t, *J* = 2.4 Hz, 1H), 4.23 (d, *J* = 12.8 Hz, 1H), 4.19 (d, *J* = 11.2 Hz, 1H), 3.95 (d, *J* = 12.8 Hz, 1H), 3.83 (dd, *J* = 11.2, 2.4 Hz, 1H), 3.62 (dd, *J* = 10.8, 7.2 Hz, 1H), 3.48 (d, *J* = 11.2 Hz, 1H), 3.39 (dd, *J* = 11.2, 4.4 Hz, 1H), 2.19–2.05 (m, 3H), 1.96 (t, *J* = 14.8 Hz, 1H), 1.82–1.74 (m, 2H), 1.38–1.29 (m, 2H), 1.20 (s, 3H), 0.79 (s, 3H) ppm; ¹³C NMR (100 MHz, CD₃OD): δ 138.7, 126.3, 81.4, 66.8, 65.0, 61.3, 55.9, 51.7, 43.1, 38.8, 36.5, 28.8, 24.4, 23.3, 16.1 ppm; HRMS (ESI): calcd for C₁₅H₂₆O₄Na [M + Na]⁺ 293.1723; found 293.1727. Compound **10**: *R*_f = 0.55 (CHCl₃/CH₃OH = 10:1); [α]_D²⁰ –81 (c 1.2, CH₃OH); IR (film): ν_{max} 3340, 2937, 2858, 1700, 1448, 1359, 1017 cm⁻¹; ¹H NMR (400 MHz, CD₃OD): δ 4.20 (d, *J* = 12.0 Hz, 2.0H), 4.13 (d, *J* = 11.2 Hz, 1H), 4.02 (d, *J* = 11.6 Hz, 1H), 3.94 (d, *J* = 12.4 Hz, 1H), 3.41 (d, *J* = 11.2 Hz, 1H), 3.36 (dd, *J* = 10.8, 5.6 Hz, 1H), 2.35 (dd, *J* = 18.0, 5.6 Hz, 1H), 2.12 (ddd, *J* = 18.4, 11.6, 7.2 Hz, 1H), 1.95–1.77 (m, 4H), 1.51–1.42 (m, 2H), 1.22 (s, 3H), 1.23–1.21 (m, 1H), 1.00 (s, 3 H) ppm; ¹³C NMR (100 MHz, CD₃OD): δ 144.3, 136.8, 81.1, 65.2, 63.2, 57.6, 52.9, 43.7, 39.0, 35.5, 31.2, 28.9, 23.4, 21.6, 20.1 ppm; HRMS (ESI): calcd for C₁₅H₃₀NO₄ [M + NH₄]⁺ 288.2169; found 288.2173.

Synthesis of (+)-Iresin (4). A mixture of **9** (57 mg, 0.21 mmol) and silver carbonate–Celite (0.57 g/mmol, 600 mg, 1.05 mmol; Fétizon reagent was prepared according to the literature methods^{15d}) in 3 mL of benzene was heated to reflux for 5 h, cooled, and filtered through a pad of Celite (ethyl acetate). The solvent was removed in vacuo, and the residue was then purified by silica gel column chromatography (petroleum ether/EtOAc = 1:1) to give (+)-iresin **1** (44 mg, 78%) as colorless crystals and aldehyde **11** (8 mg, 14%) as a white solid. NaBH₄ (2 mg, 0.052 mmol) at 0 °C was added to a solution of aldehyde in CH₃OH (1 mL). After stirring for 10 min at 0 °C, the reaction mixture was diluted with water (3.0 mL) and extracted with ethyl acetate (3 × 10 mL). The combined organic extracts were washed with 1 N aq HCl, water, and brine, dried over anhydrous Na₂SO₄, and concentrated in vacuo. The residue was purified by flash silica gel column chromatography (petroleum ether/EtOAc = 1:1) to give iresin (7 mg, 86% from **11**) as white solids. Iresin (**4**): *R*_f = 0.47 (AcOEt); mp 139–142 °C, (Lit.⁵ mp 140–142 °C); [α]_D²⁰ +20 (c 1.0, CHCl₃), (Lit.⁵ [α]_D²⁸ +21); IR (film): ν_{max} 3383, 2931, 2870, 1755, 1687, 1423, 1221, 1028, 964 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 6.86 (ddd, *J* = 3.6, 3.6, 3.6 Hz, 1H), 4.40 (dd, *J* = 9.2, 9.2 Hz, 1H), 4.26 (d, *J* = 10.4 Hz, 1H), 4.02 (dd, *J* = 9.2, 9.2 Hz, 1H), 3.54–3.49 (m, 2H), 3.12 (dd, *J* = 8.8, 2.0 Hz, 1H), 3.00 (d, *J* = 4.4 Hz, 1H), 2.83–2.77 (m, 1H), 2.51(ddd, *J* = 20.0, 4.8, 4.0 Hz, 1H), 2.18–2.10 (m, 1H), 1.89–1.79 (m, 2H), 1.68 (ddd, *J* = 13.6, 3.6, 3.6 Hz, 1H), 1.50 (dd, *J* = 12.0, 5.2 Hz, 1H), 1.35 (ddd, *J* = 13.6, 13.6, 4.0 Hz, 1H), 1.28 (s, 3H), 0.77 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 169.8, 135.8, 127.0, 80.4, 67.2, 63.3, 50.6, 50.0, 42.1, 37.1, 33.7, 27.3, 24.6, 22.1, 14.3 ppm; HRMS (ESI): calcd for C₁₅H₂₃O₄ [M

+ H]⁺ 267.1591; found 267.1585. Aldehyde **11**: $R_f = 0.64$ (AcOEt); mp 145–147 °C; $[\alpha]_D^{20} +35$ (c 0.6, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 10.00 (d, $J = 1.2$ Hz, 1H), 6.86 (d, $J = 3.2$ Hz, 1H), 4.44 (dd, $J = 8.8, 8.8$ Hz, 1H), 4.03 (dd, $J = 8.8, 8.8$ Hz, 1H), 3.37 (br d, $J = 7.6$ Hz, 1H), 2.85–2.80 (m, 2H), 2.67 (dddd, $J = 20.4, 5.2, 4.0, 4.0$ Hz, 1H), 2.49–2.45 (m, 1H), 1.97–1.94 (m, 2H), 1.75 (ddd, $J = 13.6, 4.4, 4.4$ Hz, 1H), 1.66 (dd, $J = 11.6, 6.0$ Hz, 1H), 1.42–1.35 (m, 1H), 1.34 (s, 3H), 0.74 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 206.4, 169.5, 134.6, 127.1, 76.9, 67.2, 51.9, 50.6, 49.4, 37.3, 34.2, 27.7, 24.5, 19.6, 13.6 ppm.

Synthesis of (–)-Isoiresin (13). Lactonization of **10** to aldehyde **12** (91% yield) was carried out by a procedure analogous to that of **9**. Compound **12**: white solids; $R_f = 0.57$ (AcOEt); mp 151–154 °C; $[\alpha]_D^{20} -67$ (c 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 9.83 (d, $J = 2.4$ Hz, 1H), 4.77–4.65 (m, 2H), 3.29 (br s, 1H), 3.16 (d, $J = 10.0$ Hz, 1H), 2.52–2.47 (m, 1H), 2.29–2.15 (m, 2H), 2.07–1.95 (m, 2H), 1.86–1.80 (m, 1H), 1.76 (ddd, $J = 13.2, 3.6, 3.6$ Hz, 1H), 1.55 (dd, $J = 12.8, 4.4$ Hz, 1H), 1.47 (d, $J = 12.0$ Hz, 1H), 1.37 (s, 3H), 1.09 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 206.9, 173.7, 167.9, 124.1, 77.0, 68.3, 52.5, 51.5, 36.4, 34.0, 27.8, 21.5, 20.1, 19.2, 18.2 ppm.

Reduction of aldehyde **12** to isoiresin (**13**) (92% yield) was carried out by a procedure analogous to that of aldehyde **11**. Isoiresin (**13**): colorless crystals; $R_f = 0.46$ (AcOEt); mp 217–220 °C; $[\alpha]_D^{20} -62$ (c 1.0, CHCl₃); IR (KBr): ν_{\max} 3303, 2924, 2853, 1736, 1670, 1643, 1430, 1384, 1196, 1011, 989 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 4.76–4.62 (m, 2H), 4.23 (dd, $J = 11.2, 2.0$ Hz, 1H), 3.54–3.49 (m, 1H), 3.41 (t, $J = 9.6$ Hz, 1H), 2.72 (dd, $J = 8.8, 2.4$ Hz, 1H), 2.61 (d, $J = 4.0$ Hz, 1H), 2.43 (dt, $J = 18.0, 2.8$ Hz, 1H), 2.17–2.10 (m, 1H), 2.03–1.86 (m, 3H), 1.73 (ddd, $J = 12.8, 3.2, 3.2$ Hz, 1H), 1.53–1.45 (m, 2H), 1.31 (s, 3H), 1.14 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 174.0, 169.2, 123.8, 80.1, 68.1, 63.8, 51.0, 42.9, 36.0, 33.8, 27.5, 22.6, 21.7, 21.5, 18.1 ppm; HRMS (ESI): calcd for C₁₅H₂₃O₄ [M + H]⁺ 267.1591; found 267.1586. X-ray crystallographic data of **13**: C₁₅H₂₂O₄, orthorhombic, space group: C222₁, $a = 6.580$ (6) Å, $b = 12.892$ (15) Å, $c = 32.76$ (3) Å, $Z = 8$, $d_{\text{calcd}} = 1.359$ g/cm³, $R_1(I > 2\sigma(I)) = 0.0357$, $wR_2 = 0.0836$.

Preparation of Bis(4-Br)benzoate Derivative 14 of (+)-Iresin (4). To a stirred solution of (+)-iresin (**4**) (43 mg, 0.16 mmol) in dry pyridine (2 mL) were added *p*-bromobenzoyl chloride (177 mg, 0.81 mmol) and DMAP (2 mg, 0.016 mmol) at room temperature. When the consumption of iresein was complete (monitored by TLC), the reaction mixture was extracted with ethyl acetate, then washed with saturated NaHCO₃ solution, water, and brine, dried over Na₂SO₄, and concentrated to give a light yellow solid residue, which, after purification by flash column chromatography on silica gel (benzene/EtOAc = 30:1), afforded **14** (97 mg, 95%) as colorless crystals. $R_f = 0.30$ (petroleum ether/EtOAc = 1:2); mp 208–211 °C, (Lit.^{6e} mp 211.5–212 °C); $[\alpha]_D^{20} -50$ (c 1.6, CHCl₃); IR (KBr): ν_{\max} 3014, 2890, 1760, 1741, 1689, 1590, 1480, 1291, 1012, 846 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 7.81–7.78 (m, 4H), 7.53 (d, $J = 8.4$ Hz, 2H), 7.40 (d, $J = 8.4$ Hz, 2H), 6.90 (d, $J = 3.2$ Hz, 1H), 4.96 (dd, $J = 9.6, 6.4$ Hz, 1H), 4.87 (d, $J = 12.0$ Hz, 1H), 4.59 (d, $J = 11.6$ Hz, 1H), 4.45 (dd, $J = 9.2, 9.2$ Hz, 1H), 4.08 (dd, $J = 9.2, 9.2$ Hz, 1H), 2.92–2.87 (m, 1H), 2.63 (dd, $J = 20.0, 4.4$ Hz, 1H), 2.53–2.45 (m, 1H), 1.96–1.91 (m, 2H), 1.81–1.75 (m, 2H), 1.60–1.52 (m, 1H), 1.23 (s, 3H), 0.92 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 169.4, 165.8, 165.3, 135.1, 131.9, 131.6, 131.1, 130.9, 128.8, 128.7, 128.4, 128.3, 127.0, 80.2, 66.9, 64.1, 50.3, 50.2, 41.7, 36.9, 33.9, 25.1, 23.7, 22.1, 13.8 ppm; HRMS (ESI): calcd for C₂₉H₂₉Br₂O₆ [M + H]⁺ 633.0305; found 633.0309. X-ray crystallographic data of **16**:^{6e} C₂₉H₂₈Br₂O₆, monoclinic, space group: P2₁, $a = 6.2860$ (12) Å, $b = 7.3672$ (15) Å, $c = 28.272$ (6) Å, $\beta = 92.047$ (13)°, $Z = 2$, $d_{\text{calcd}} = 1.605$ g/cm³, $R_1(I > 2\sigma(I)) = 0.0530$, $wR_2 = 0.1339$.

ASSOCIATED CONTENT

Supporting Information

¹H and ¹³C NMR spectra for all compounds **4–6**, **7a**, **7b**, **8–14**, X-ray crystallographic data for compounds **6**, **13**, **14**, and CIF files for compounds **6**, **13**, **14**. The Supporting Information

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Notes

The authors declare no competing financial interest.

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DEDICATION

In memory of Carl Djerassi.

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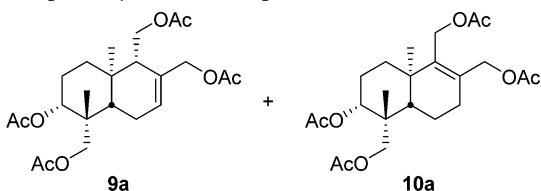
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